

Detecting invasive species with the eDNA method

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First application of eDNA to map alien, invasive species in Greek freshwaters (project RESILIENT)

Target species: Gambusia holbrooki





OBJECTIVE

Mapping the alien invasive Eastern mosquitofish *G. holbrooki* in Valencia habitats, *using BOTH c*onventional fish sampling methods and eDNA sampling







G. holbrooki

Fishing detection



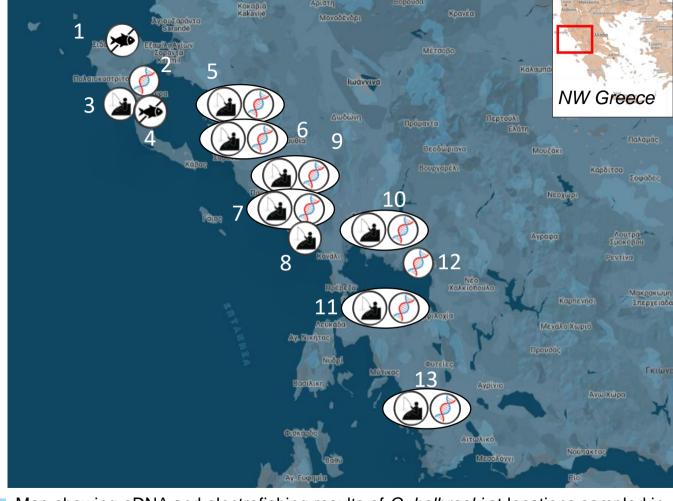
e DNA detection



No detection







Map showing eDNA and electrofishing results of *G. holbrooki* at locations sampled in W. Greece (*V. Letourneuxi* distributional range) during the 2018 autumn survey



G. holbrooki

Fishing detection



e DNA detection



No detection



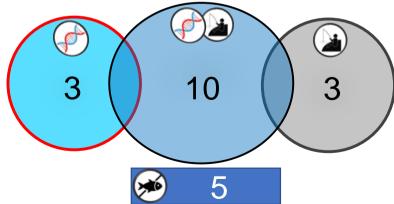


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Map showing eDNA and electrofishing results of *G. holbrooki* at locations sampled in W. Greece (*V. robertae* distributional range) during the 2018 autumn survey

Conclusions

- At 3 systems, G.holbrooki was detected through eDNA but NOT through fish sampling, → low densities, indicating the suitability of the eDNA method for species detection
- At 3 sites, G. holbrooki <u>was detected through fish sampling</u> but <u>NOT</u> through eDNA (pseudonegatives), → limitations of the method (fast flow and/or turbidity)



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Targeting two top invaders/<u>nation-wide</u> survey (project PACIM)

Target species: Gambusia holbrooki and Pseudorasbora parva





OBJECTIVE

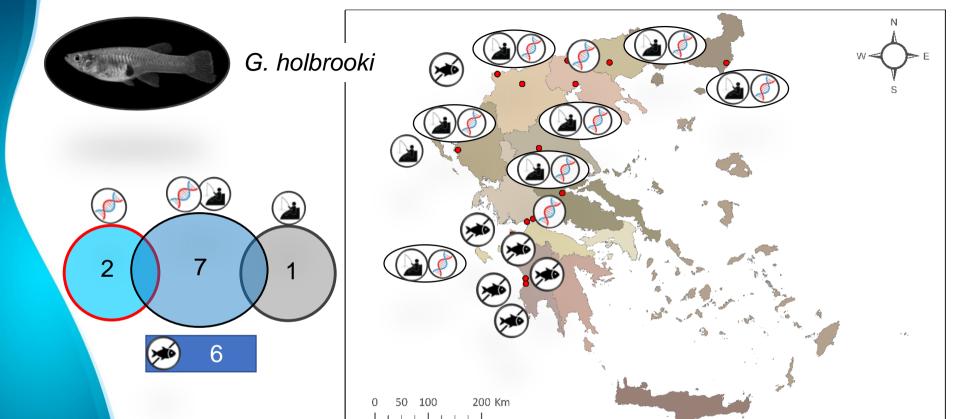
Nation-wide survey targeting two top freshwater fish invaders, using BOTH conventional fish sampling methods and eDNA sampling











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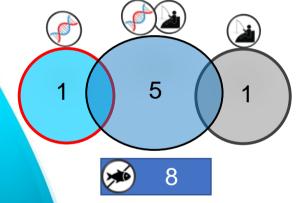


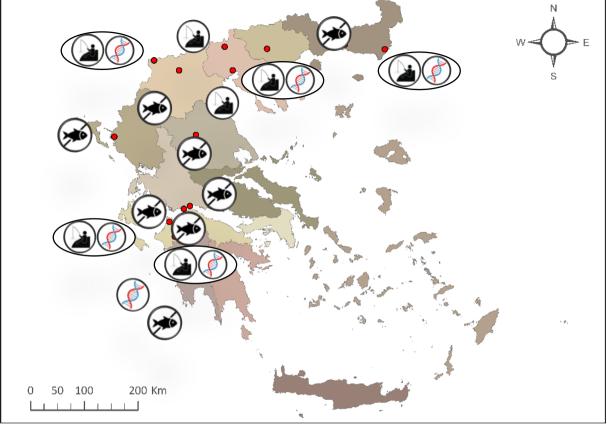
e DNA detection () Fishing detection () No detection





P. parva





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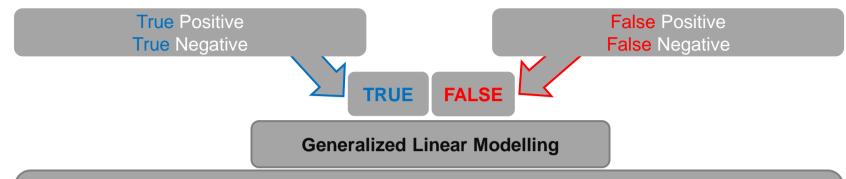
Conclusions

- At two systems (within known range), G.holbrooki was detected through eDNA but NOT through fish sampling, i.e. low densities, indicating the suitability of the eDNA method for species detection
- No eDNA pseudo-negatives for G. holbrooki

- At one system, P. parva was detected through eDNA but NOT through fish sampling (early detection of recent expansion?)
- Two eDNA pseudo-negatives for P. parva



Occupancy Modelling



23 environmental, habitat and biological variables from 16 sampling locations were regressed against the outcome of the eDNA analysis in order to identify the most significant predictors

- 1. Fish species
- Filtered Volume
- 3. Ecoregion
- 4. Latitude
- 5. Longitude
- 6. Temperature

- 7. pH
- 8. Turbidity
- 9. D.O. Saturation
- 10. Flow
- 11. Fast habitat %
- 12. Wetted Width

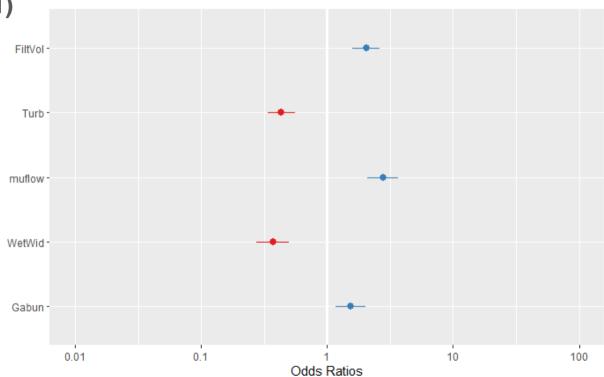
- 13. Depth
- 14. Coarse Substrate %
- 15. Shadedness
- 16. Helophytes
- 17. Bottom Vegetation
- 18. G. hol. abundance

- 19. G.hol. density
- 20. G. hol. abundance/Min
- 21. P. par. abundance
- 22. P. par. density
- 23. P. par abundance/Min



RESULTS Effect of Standardized Coefficients

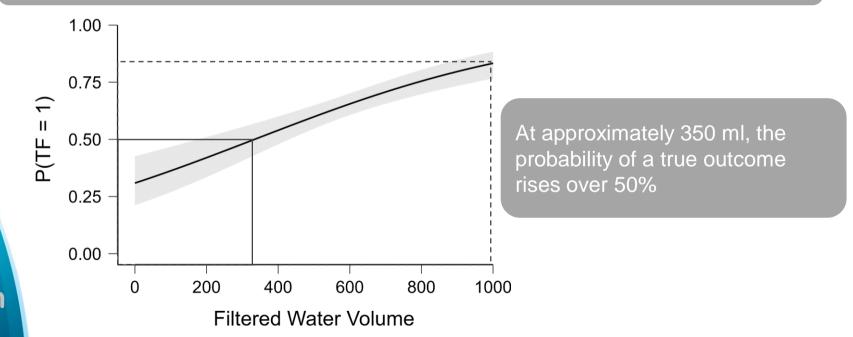




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Conditional Plot of Filtered Water Volume

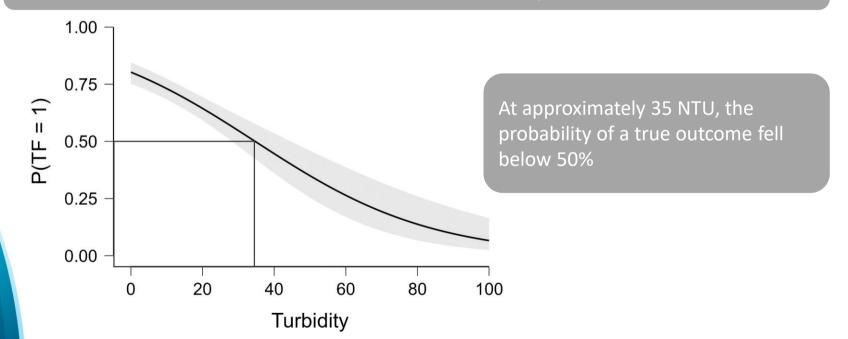
The odds of a TRUE replicate outcome occurring increased by 0.24% for every unit of increase in the Volume of Filtered Water per replicate sample.





Conditional Plot of Turbidity

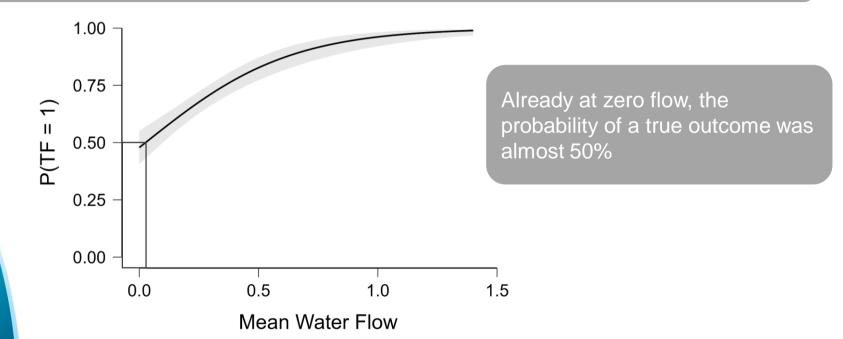
The odds of a TRUE replicate outcome occurring decreased by 3.97% for every unit of increase in water turbidity.



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Conditional Plot of Mean Water Flow

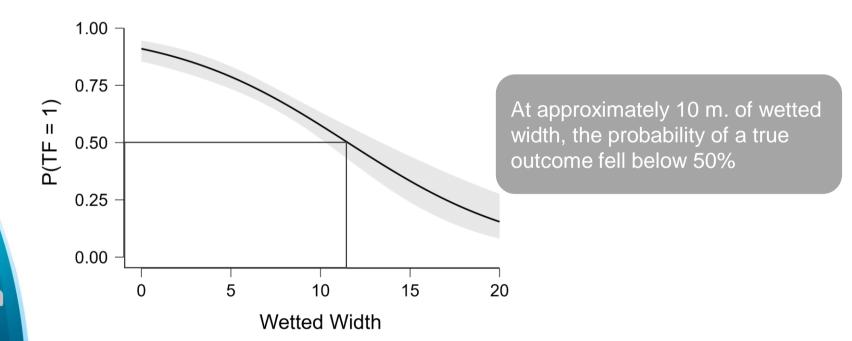
The odds of a TRUE replicate outcome occurring increased by 2618.91% for every unit of increase in water flow.



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Conditional Plot of Wetted Width

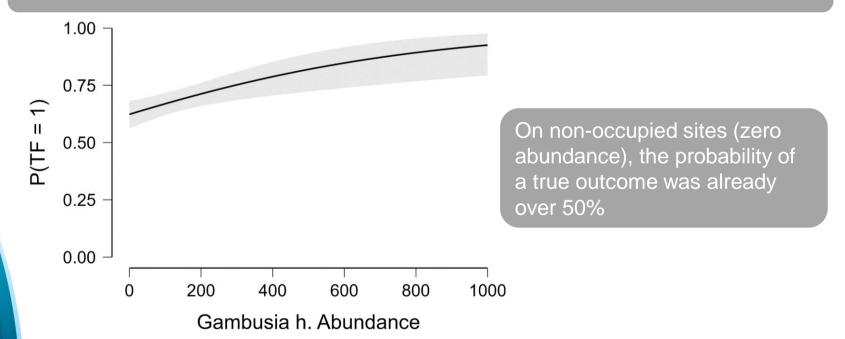
The odds of a TRUE replicate outcome occurring decreased by 18.18% for every unit of increase at wetted width.



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Conditional Plot of G. holbrooki Abundance

The odds of a TRUE replicate outcome occurring increased by 0.2% for every unit of increase in the abundance of *G. holbrooki*



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Widening the scope, two more top invaders/<u>nation-wide</u> survey in Greek freshwaters (project AFRESH)

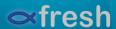
Target species: Carassius gibelio and Lepomis gibbosus





OBJECTIVE

Nation-wide survey targeting two more top freshwater fish invaders, Using BOTH conventional fish sampling methods and eDNA sampling





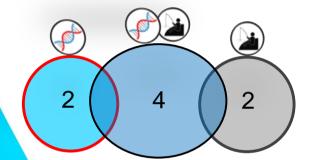






C. gibelio

15 Evros



C. gibellio **River Basins** Test type eDNA Fishing Negative 1 Assopos Beot. Test Negative 2 Assopos Beot. Test Negative Negative 3 Kifissos Beot. Test Negative Negative 4 Pinios Thes. Positive Positive Test 5 Pinios Thes. Test Negative Positive 6 Prespa (Ag. Germanos) Control Negative Negative 7 Aliakmon Test Negative Negative Positive 8 Axios Test Negative Positive 9 Doiran Test Positive 10 Strymon Test Negative Negative 11 Nestos Control Negative Negative Test Positive Negative 12 Kossynthos 13 Filiouris Test Positive Positive 14 Evros Test Negative Positive Test Positive Positive

Βόρεια Μακεδον

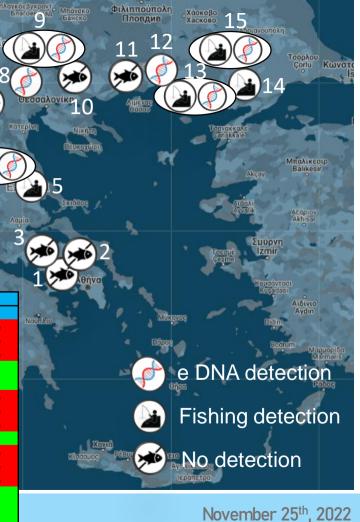
Мпітола

Tipava Tiranë

AXBavla 6

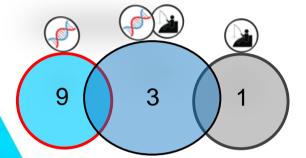
Aylo Zapavta Saranda





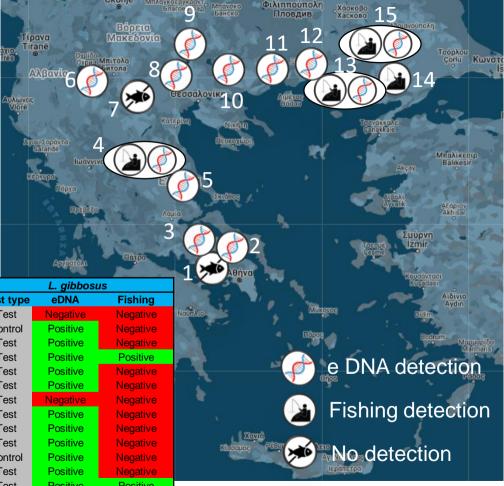


L. gibbosus



6 2

	L. gibbosus		
River Basins	Test type	eDNA	Fishing
1 Assopos Beot.	Test	Negative	Negative
2 Assopos Beot.	Control	Positive	Negative
3 Kifissos Beot.	Test	Positive	Negative
4 Pinios Thes.	Test	Positive	Positive
5 Pinios Thes.	Test	Positive	Negative
6 Prespa (Ag. Germanos)	Test	Positive	Negative
7 Aliakmon	Test	Negative	Negative
8 Axios	Test	Positive	Negative
9 Doiran	Test	Positive	Negative
10 Strymon	Test	Positive	Negative
11 Nestos	Control	Positive	Negative
12 Kossynthos	Test	Positive	Negative
13 <u>Filiouris</u>	Test	Positive	Positive
14 Evros	Test	Negative	Positive
15 Evros	Test	Positive	Positive



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e-Workshop

November 25th, 2022

Conclusions

- At 2 systems (within known range), *C. gibelio* was detected through
 eDNA but NOT through fish sampling, i.e. low densities, indicating the suitability of the eDNA method for species detection
- One eDNA pseudo-negative for C. gibelio

- At 5/9 systems, L. gibbosus was detected through eDNA but NOT through fish sampling (early detection of recent expansion?)
- One eDNA pseudo-negatives for L. gibbosus!





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e-Workshop

November 25th, 2022