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# Advantages and Shortcomings of the eDNA method

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afresh

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# Short introduction to eDNA methods

Looking for traces of target species' DNA in the environment

Environmental DNA can be found in:



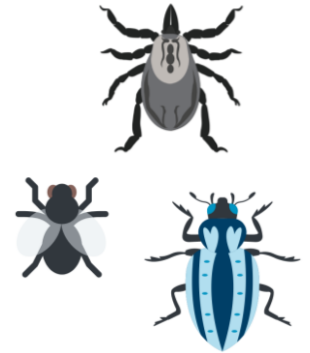
Water



Surface soil and  
soil/sediment cores



Air and  
cavities



Invertebrates  
(iDNA)

# Why eDNA?

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## General advantages of eDNA methods

- **Non-invasive:** The collection does not require contact with the animal, which makes it a lot more suitable for rare or elusive species.
- **Large coverage:** Some sample types have a large monitoring range, allowing us to cover challenging terrain and areas with no access.
- **Minimised surveyor bias:** Sample collection is not dependent on ID skills.
- **Survey-specific advantages / species-specific advantages**

# eDNA methods: the technical part

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## Fieldwork

Sampling techniques vary across the different sample types, but they all have the following in common:

- **Low budget** field equipment can be used.
- **Minimal training** is required for sample collection.
- **Short fieldwork time** in comparison with other techniques.
- Samples are **preserved** until lab processing.

# eDNA methods: the technical part

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## Lab work

Laboratory analysis depends on what we are looking for in our samples:

Single-species, or a small selection of known species



**Species-specific or multiplexing**

Species diversity in the sample



**Metabarcoding**

# Species-specific versus metabarcoding

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What are the benefits of each method?

## Species-specific

More sensitive for rare species

Analysis costs are low

Less reliant on hi-spec equipment

## Metabarcoding

Can miss rarer DNA in the sample

Analysis costs are high

Relies on highly trained lab techs and metabarcoding analysts

# What can eDNA data tell us?

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## Using eDNA survey results to gather information on a species

eDNA gathers information on **species presence**.

Work is currently being done for most survey methods to define **what a 'detection' means**. How long has DNA persisted in the medium? How far has the DNA sample travelled? Does the amount of DNA in the sample equate with population size/activity patterns?

Positive detections can be used **for modelling the species' distribution** (MaxEnt)

For repeat surveys, data (positive and negative detections) can be used to run **occupancy modelling**.

# Weaknesses of eDNA methods

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**eDNA methods are new and research is currently focusing on advancements and refinement of the different methods**

- On single-species methods, the laboratory analysis needs to be adjusted to each species (primer/probe design), which **requires expertise**.
- eDNA samples **processing time** is long in comparison to direct results from other field survey methods.
- Any molecular lab work can be subject to **contamination**, often minimised by repeat sampling.



# What my results showed

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## Advice from my personal experience with eDNA sampling

As we adopt eDNA on our monitoring efforts there are a few things that will **increase the reliability** of the results:

- **Repeat sampling** (sampling multiple times, over multiple seasons)
- **Repeat extractions** DNA of each sample
- **Amplification replicates** for each extracted sample

Repetition in these three survey stages will ensure the accuracy of the results and help us gather a vast amount of information for the species' ecology and habitat use.

# Next steps with eDNA

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## eDNA has truly advanced species monitoring

Research is currently focusing on:

- Developing more eDNA **sampling techniques**
- Understanding what '**positive detection**' means for each method
- **Refining** the laboratory methods
- Designing **species-specific** methods for single-species detection
- Use of eDNA data in **spatial modelling**
- **Widening** the use of eDNA methods and **lowering** the processing costs

# Thank you

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Thank you to Dr Mark Steer, my PhD supervisor

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